

Investigation into the Pharmacognostic and Antimicrobial Properties of the Leaf Extract of *Bridelia Ferruginea* Benth (*Euphorbiaceae*)

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ABSTRACT

Introduction:

Bridelia ferruginea is a well-known shrub that is used in traditional medicine in the management of certain diseases such as dysentery, arthritis and constipation.

Aim/objectives: to ascertain the validity of the use of *B. ferruginea* as an antimicrobial agent, to evaluate the pharmacogenetic profile and differentiate it from adulterated one, to detect the secondary metabolites present in the leaf extract and to compare the plants antimicrobial activity with standard antimicrobial agents.

Methods: The leaves of *Bridelia ferruginea* were plucked from the stem and dried in the shade for approximately 1 week. The leaves were ground into moderately coarse powder using mortar and pestle. A 400 g quantity of the grounded leaves were packed into a glass jar and macerated with methanol for 24 hours. The extracts were then filtered, distilled and concentrated in a rotary evaporator at optimum temperature of 40.5°C and was brought to complete dryness over a water bath. The flask containing the extract was allowed to cool and the extract was then collected.

Results: The MIC values of the *Bridelia ferruginea* extract against susceptible organisms (*S. aureus*, *B. subtilis* and *C. albicans*) recorded results of 9.5 ± 0.21 mm, 7.0 ± 0.15 mm and 9.0 ± 0.20 mm respectively without any significant difference ($p < 0.05$). The values of the inhibitory zone diameter obtained ranged from 7.0 ± 0.01 mm to 9.5 ± 0.02 mm for

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C. albicans and *S. aureus* respectively, without any significant difference ($p < 0.05$). From the results obtained, the extract was more effective only against Gram positive bacteria. The phytochemical analysis of the leaves of *Bridelia ferruginea* showed the presence of secondary metabolites.

Conclusion: The study indicates the enormous potential of *B. ferruginea* with anti-microbial properties. Due to its wide application, the phytochemicals present has been documented, and they could be useful in lead compound discovery.

Keywords: *Bridelia ferruginea*, Phytochemicals, Metabolites, Anti-microbials, Extract.

INTRODUCTION

Over the years, the knowledge and practice of traditional medicine has majorly influenced drug development [1]. Majority of the conventional medicines used all over the world originated from plants or its related compounds. Drug companies have leveraged on this advantage by the continuous development of herbs into conventional medicine [2]. A lot of disease management has now resorted to the use of medicinal plants and herbs especially in developing countries like Nigeria. The reason for such has been attributed

to inaccessibility to conventional healthcare, cost and cultural acceptability [3].

Bridelia ferruginea is a well-known medicinal plant that belongs to the Euphorbiaceae family. Its pharmacological properties have been documented [4]. Some of the plant parts like fruits, leaves, barks and roots have been commonly used as decoctions, in arthritis management, dysentery, constipation, diarrhea, skin and intestinal infections [5]. According to Aja et al, 2013, [6] the antimicrobial, repellent and fibroblast growth stimulation properties of the leaves of *B. ferruginea* has been documented. The stem and stem bark have demonstrated anti-inflammatory, antioxidant, antimicrobial, antityphoid, anthelmintic and analgesic properties [7-15].

The antimicrobial activities of leaf extracts of *B. ferruginea* against *Staphylococcus aureus*, *Shigella dysenteriae* and *Escherichia coli*, and *Bacillus subtilis* has been reported by Fadeyi and Akpan [16]. *Bridelia Ferruginea* is a shrub that is commonly found in fields especially in the Northern region axis and some African countries like Congo, East Africa, Angola and Zimbabwe. There is limited information on the leaf extract of *B. ferruginea* and its antimicrobial properties [17-20].



Figure 1. A) Matured leaves before reproduction, B) New leaves [20].

MATERIALS AND METHOD

Materials

Bridelia ferruginea leaves were collected from Obukpa Nsukka in Enugu state, Dimethylsulphoxide (DMSO) (Sigma-Aldrich), Nutrient agar (Oxoid Ltd, England), sterile distilled water, cotton wool, indelible marker (UNN), Sabouraud Dextrose

Agar, standard antibiotic (Gentamycin), Methanol 95%, chloroform, ammonia solution, ethanol, Picric acid solution (Oxoid Ltd, England), lead acetate solution, Wagner's reagent -Mayer's reagent Dragendorff, ferric chloride solution, dilute hydrochloric acid solution (Oxoid Ltd, England), glacial acetic acid, dilute sulphuric acid, sodium carbonate solution, 10% aqueous aluminium chloride solution (Oxoid Ltd, England),

Bromine water, Gram positive bacteria: Staphylococcus aureus Bacillus subtilis Gram negative bacteria: Escherichia coli, Pseudomonas aeruginosa, Klebsiella pneumoniae, Salmonella typhi Fungi Yeast (Candida albicans), Mould (Aspergillus niger) (Gattefosse Saint-Priest Cedex, France). All other reagents and solvents used were of analytical grade.

Method

Sourcing of the leaves of *B. ferruginea*

Bridelia ferruginea leaves were collected from Obukpa Nsukka in Enugu state and identified by a taxonomist, Mr. Felix Nwafor, a Botanist, in the Pharmacognosy Herbarium, University of Nigeria Nsukka, Enugu State Nigeria. A herbarium Voucher Number of PCG/UNN/04980 Euphorbiaceae (L) was assigned.

Extraction procedure

The leaves of *Bridelia ferruginea* were plucked from the stem and dried in the shade for approximately 1 week. The leaves were ground into moderately coarse powder using mortar and pestle. A 400 g quantity of the grounded leaves were packed into a glass jar and macerated with methanol for 24 hours. The extractives were then filtered, distilled and concentrated in a rotary evaporator at optimum temperature of 40.5°C and was brought to complete dryness over a water bath. The flask containing the extract was allowed to cool and the extract was then collected [21-23].

Antimicrobial evaluation of the methanolic extract of *B. ferruginea*

Sterilization of materials

The pipettes, test tubes and the agar plates packed into metal canisters were sterilized using hot air oven at 170°C for 1 hour. The oven was put off and the metal canisters brought out and allowed to cool before the materials inside were brought out for use. The culture media were sterilized using autoclave at 121°C for 15 minutes.

Preparation of test microorganisms

The test organism's suspensions were prepared by marching the turbid suspension with a Marcfarland standard 0.5. Stock solutions of microorganisms were subcultured and incubated at 37°C and 25°C for bacteria and fungi respectively for 18-24 hours before use, in order to get pure isolates.

Standard suspensions of each test microorganism were made by transferring a colony from the subculture into 5 ml of sterile distilled water and adjusting the volume to obtain a cell population of 1.0×10^6 cfu/ml which was the standard inoculum size used in all the tests [24].

Preparation of culture media

In the preparation of sabouraud dextrose medium for screening of plant extract using fungi, 0.05mg/ml of chloramphenicol was incorporated into the medium to preclude interference or growth of bacteria in the fungi medium. A 250 mg quantity of chloramphenicol was dissolved in 5 ml of ethanol, then 1 ml was withdrawn from the solution and then made up to 100 ml of the sabouraud dextrose medium using distilled water. A 65 g quantity of sabouraud dextrose powder leaf was suspended in 1 litre of distilled water containing 0.05mg/ml chloramphenicol and it was soaked for 5 minutes until all the powder has dissolved. The mixture was swirled, and sterilized by autoclaving at 121°C for 15 minutes, cooled to 48- 50°C and then poured into a petri dish [25].

Preparation of Nutrient agar medium for bacteria Formulation

Nutrient agar Peptone 5.0 g, Beef extract 3.0 g, Sodium chloride 8.0 g, Agar no 2 12.0 g pH 7.3 \pm 2.28g of Nutrient agar powder was dispersed in 1 litre of deionized water and the mixture was soaked for 10 minutes. It was swirled to mix properly and then sterilized for 15 minutes at 121 oC. It was cooled to 47 oC, poured into a plate and stored away from direct sunlight [26].

Preparation of stock solution of plant extract

The stock solution of methanolic extract of *Bridelia ferruginea* (100 mg) was prepared by weighing 200 mg of the substance and dissolving some in 2ml of dimethylsulfoxide (DMSO) and agitating to complete dissolution. This was used as the stock solution, serial dilutions of the extract were obtained: 100mg/ml, 50mg/ml, 25mg/ml, 12.5mg/ml and 6.25mg/ml [26].

Preliminary antimicrobial screening (sensitivity test)

Preliminary antimicrobial screening of the methanolic extract was done using the cup plate agar diffusion method. A 0.2 ml portion of each test organism was added to 20 ml of sterile molten agar (bacteria and fungi media) in seven

different nutrient agar plates, mixed thoroughly and allowed to equilibrate [27]. The seeded agar plates were allowed to set for about 10 minutes. A sterile cork borer, 8 mm in diameter, was then used to form cups at five different points in each plate. Each plate was inoculated with a microorganism and labeled appropriately. Afterwards, two drops of the methanolic extract, dissolved in dimethyl sulfoxide were introduced into each cup using sterile Pasteur pipettes. The plates were then allowed to stand for 30 minutes at room temperature to allow for pre-diffusion and then incubated at 37°C for 24 hours. After incubation, the diameters of the zones of inhibition were measured in millimeter for the sensitive organisms [28,29].

Determination of minimum inhibitory concentration (MIC)

The MICs of methanolic extracts of the plant were determined against susceptible organisms using agar diffusion method [30]. Agar plates were prepared (seeded) with organisms in two different plates; Graded concentrations of the extracts (100mg/ml, 50 mg/ml, 25 mg/ml, 12.5 mg/ml, 6.25 mg/ml) were prepared. Six wells were bored on seeded plates using a sterile cork borer of 8 mm diameter. Each well was marked and 1ml volumes of the different dilutions introduced into them using a sterile pipette. The test was performed for both extracts using each test organism and each in duplicate and the plates were incubated at 37°C for 24 hours (for bacteria) and 25°C for 48 hours (for fungi). The inhibitory zone diameters were recorded by calculating the mean of each set of replicate inhibition zone diameters. The minimum inhibitory concentrations (MICs) were determined by plotting a graph of inhibitory zone diameters (IZD) against the logarithm of the concentrations. The above procedure was also carried out to determine the MIC of Gentamycin and Clotrimazole against the sensitive bacteria and fungi respectively.

Phytochemical analysis

Phytochemical analysis for crude drug includes:

Test for Resins

A 0.3 g of the crude drug was extracted with 10 ml of 96 % ethanol. The alcoholic extract was then poured into 20 ml of distilled water in a beaker and observed [31].

(b) Colour test: Another 0.3 g quantity of the crude drug was extracted with chloroform and the extract was concentrated

to dryness. The residue was concentrated to dryness. The residue was redissolved in 3 ml of acetone and 3 ml of concentrated hydrochloric acid was added. The mixture was heated in a water bath for 30 minutes and observed.

Test for Alkaloids

A 0.5 g quantity of the extract was warmed on a water bath with 2 ml of 1% Sulphuric acid for 2 minutes. It was filtered and the filtrate used for the following tests;

- a. Mayer's test 2 drops of Mayer's (Potassium mercuric iodide) solution was added to 5ml of the filtrate. A creamy of white buff precipitate indicates the presence of alkaloid [32].
- b. Dragendorff's test: A 2 drops quantity of Dragendorff's reagent (potassium bismuth iodide) solution was added to 5ml of filtrate. A deep brick-red precipitate indicates the presence an alkaloid [33].

Test for Tannins

A 1g quantity of the methanolic extract of the leaves was boiled with 20 ml of water, cooled and filtered. The filtrate was then used for the following tests:

- a. Ferric chloride test: A 1 ml quantity of the filtrate was diluted to 5 ml with water and a few drops of ferric chloride (0.1%) were added. A greenish brown precipitate indicates the presence of tannins.
- b. Bromine water test: A 1ml quantity of the filtrate was added 2 drops of bromine water. A pale brown precipitate indicates the presence of tannins [34].

Test for Flavonoids

A 0.2 g quantity of the methanolic extract was heated with 10 ml of ethyl acetate in a boiling water bath for 3 minutes. The mixture was filtered and the filtrate was used for the following tests;

- a. Ammonia test: A 4 ml quantity of the filtrate was shaken with 1ml of dilute ammonia. A yellow coloration in ammonia layer indicates the presence of flavonoids.
- b. Aluminium chloride test: A 4 ml quantity of filtrate was shaken with 1 ml of 10 % Aluminium chloride solution. A yellow colour in the aluminium layer indicates the presence of flavonoids [35].

Test for Saponins

A 1 g quantity of the methanolic extract of the leaves was boiled with 10 ml of distilled water for 10 minutes, filtered while hot and filtrates cooled and the following tests were performed;

a. Frothing test

A 2.5 ml quantity of the filtrate was diluted to 10ml with water and shaken vigorously for 2minutes. Persistent foaming (stable froth) indicates the presence of saponins.

b. Emulsion test

2 drops of olive oil was added to the filtrate and shaken vigorously. Formation of stable emulsion indicates the presence of saponins [36].

Test for Steroids

A 1g quantity of the methanolic extract was boiled under reflux with 18ml of ethanol and filtered. The filtrate was concentrated to about 15 ml and 10 ml of water was added to the concentrated filtrate and allowed to stand for about one hour and the waxy material filtered. The filtrate was extracted with chloroform and used for the following tests;

a. Salkowski test: A 1ml quantity of sulphuric acid was carefully added to 0.5 ml of the concentrated chloroform extract in a test tube, so as to form a lower layer and heated for 15 minutes. A red interphase between the 2 layers indicates the presence of steroids [35,36].

Test for Terpenoids

a. Liebermann's test: A 0.5 ml quantity of the concentrated chloroform extract in a test tube was evaporated to dryness. The residue was dissolved in a 0.5 ml glacial acetic acid and 1 ml sulphuric acid and heated for 15 minutes. A red magnate or violet blue indicates the presence of terpenoids [38].

Test for oils

A 0.1 g quantity of the powdered drug was pressed between filtered paper and the paper observed. Translucency of the filter paper indicates the presence of oil [39].

Test for Glycosides

A 5ml quantity of dilute sulphuric acid was added to 0.1 g of the powdered drug in a test tube and boiled for 15 minutes on a water bath, then cooled and neutralized with 20 % potassium hydroxide solution. A 10 ml quantity of a mixture of equal parts of Fehling's solution 1 and 2 were added and boiled for 5 minutes. It was then observed for the presence of brick-red precipitate [40].

Test for reducing sugars

A 0.2g quantity of the powdered drug was shaken vigorously with 10 ml of distilled water and filtered. The filtrate was used in the following tests [41]:

a) Fehling's test: To 1 ml portion of the filtrate, equal volumes of fehling's solution 1 and 11 were added and boiled on water bath for few minutes. A brick-red precipitate indicates the presence of reducing sugars.

b) Benedicts's test: To 1 ml portion of the filtrate was added 2 ml of Benedict's reagent. A rusty brown precipitate indicates the presence of reducing sugar.

Test for Protein

A 0.5 g portion of the powdered drug was extracted with 10 ml of distilled water and the filtrate was used for the following tests:

a) Million's test: To a little portion of the filtrate in a test tube, two drops of Million's reagent were added. A white precipitate indicates the presence of proteins.

b) Xanthoproteic reaction test: A 5ml quantity of the filtrate was heated with few drops of concentrated nitric acid. A yellow colour which changes to orange on addition of alkali indicates the presence of proteins [41].

c) Picric acid test: A few drops of picric acid were added to a little portion of the filtrate and observed.

Test for Carbohydrates

A 0.1 g quantity of the powdered material was boiled in 2ml of water and filtered. To the filtrate, few drops of Naphthol solution in ethanol (Molisch reagent) was added. Concentrated sulphuric acid was then gently poured down the

side of the test tube to form a lower layer. A purple interfacial ring indicates the presence of carbohydrates [42].

Test for acidity

The plant material was placed in a clean dry test tube and sufficient water was added. This was warmed in hot water bath and then cooled. A piece of water-wetted litmus paper was dipped into the filtrate and colour change on the litmus paper was observed.

Microscopic evaluation

A little quantity of the powdered drug was placed on a glass slide and a drop of chloral hydrate was added. It was covered with a cover slip and passed three times over the flame from a Bunsen burner. It was then focused on the microscope and viewed.

Standardization procedures

Total Ash value

A tarred nickel crucible was ignited to a constant weight at a dull red heat, cooled and stored in a desiccator. A 2 g quantity of the powdered material was weighed into the nickel crucible and heated gently until all the moisture had been completely removed, after which the material was heated further until residue was free from carbon. The residue was cooled in a desiccator and weighed. The heating and cooling continued until a constant weight was achieved and the total ash value calculated from the results [43-45].

Acid Insoluble Ash Value

The total ash obtained in total ash value experiment was transferred to a beaker containing 25 ml of dilute hydrochloric acid and heated to boiling on a water bath for five minutes. It was then filtered through ash-less filter paper and the beaker and crucible were washed with distilled water until free from acid. The filter paper was dried in the oven, folded into a narrow cone and inserted into a tarred nickel crucible and heated until it was completely ashed. The residual ash was then heated more strongly and after cooling in a desiccator, it was weighed [43-45].

Sulphated Ash value

A nickel crucible was ignited to a constant weight at a dull red heat. A 2g quantity of the material was spread over the bottom of the crucible and was reweighed. The material was moistened with dilute sulphuric acid and ignited at low heat initially to burn off the carbon content. The crucible was cooled in a desiccator and more dilute sulphuric acid was added and heating continued with occasional cooling until a constant weight was obtained [46].

Moisture content

A tarred evaporating dish was heated to a constant weight and stored in a desiccator. A 2g quantity of the powdered material was added and the dish was kept in an oven maintained at a temperature of 105°C and was allowed to dry until a constant weight was obtained.

Data analysis

All the measurements were repeated at least thrice and the data obtained analyzed by Student t-test and One-Way Analysis of Variance (ANOVA). Statistical analysis was performed using Statistical Product and Services Solution software (SPSS, version 22.0 Inc., Chicago IL, USA) and Excel Microsoft Office version 2012. The results were presented as mean \pm SD, and statistical differences between means considered significant at ($p < 0.05$).

RESULTS AND DISCUSSION

Antimicrobial evaluation of *Bridelia ferruginea* methanolic leaf extract

After the incubation period for the bacteria and fungi at 37°C for 24 hours and 25°C for 48 hours respectively, the following antimicrobial result for the methanolic extract and standard antimicrobial agents (Gentamycin and Clotrimazole) were obtained as shown in Table 1 below. The MIC values of the *Bridelia ferruginea* extract against susceptible organisms are shown in Table 1. *S. aureus*, *B. subtilis* and *C. albicans* recorded results of 9.5 ± 0.21 mm, 7.0 ± 0.15 mm and 9.0 ± 0.20 mm respectively without any significant difference ($p < 0.05$).

Table 1. Sensitivity of the various test organisms to the extracts of *Bridelia ferruginea*

Test organism	Result obtained (mean \pm SD)
<i>Escherichia coli</i>	++
<i>Staphylococcus aureus</i>	9.5 \pm 0.21
<i>Salmonella typhi</i>	++
<i>Pseudomonas aeruginosa</i>	++
<i>Klebsiella pneumonia</i>	++
<i>Bacillus subtilis</i>	7.0 \pm 0.15
<i>Candida albicans</i>	9.0 \pm 0.20
<i>Aspergillus niger</i>	++

Key: ++ = Presence of growth (Not sensitive)

The inhibitory zone diameter of the test organisms is represented in Table 2. The values obtained ranged from 7.0 \pm 0.01 to 9.5 \pm 0.02 for *C. albicans* and *S. aureus* respectively at 100 mm, without any significant difference ($p < 0.05$).

According to Irobi et al; the inhibitory zone diameter at 4-30mm diameter, showed that the results obtained ranged from +2 to +3 for the test organisms [47,48].

Table 2. Inhibitory zone diameters of sensitive microorganisms using different concentrations of the extract

Test organism	Average zone of inhibition (mm, mean \pm SD)				
	100	50	25	12.5	6.25
<i>Staphylococcus aureus</i>	9.5 \pm 0.02	6.0 \pm 0.02	5.3 \pm 0.01	2.0 \pm 0.01	1.8 \pm 0.01
<i>Bacillus subtilis</i>	9.0 \pm 0.02	7.5 \pm 0.02	5.0 \pm 0.01	2.0 \pm 0.01	0.8 \pm 0.01
<i>Candida albicans</i>	7.0 \pm 0.01	4.3 \pm 0.01	3.5 \pm 0.01	2.5 \pm 0.01	-

The spectrum of activity of the extract against different organisms are recorded in Table 3. The IZD of *S. aureus* ranged from 0.81 \pm 0.01 mm to 9.50 \pm 0.20 mm, while *B. subtilis*

recorded IZD values of 3.20 \pm 0.01mm to 9.00 \pm 0.03 mm. The *C. albicans* recorded IZD values of 2.50 \pm 0.01mm to 7.00 \pm 0.01mm without any significant difference ($p < 0.05$).

Table 3. Spectrum of activity of leaf methanolic extract against bacteria and fungi

Microorganism	Concentration of extract (mg/ml)	IZD = L (mm)	L ² (mm)
<i>S. aureus</i>	100.00 ± 0.23	9.50 ± 0.20	90.25 ± 0.23
	50.00 ± 0.21	7.50 ± 0.10	56.25 ± 0.20
	25.00 ± 0.10	5.00 ± 0.10	25.00 ± 0.10
	12.50 ± 0.01	2.00 ± 0.01	4.00 ± 0.10
	6.25 ± 0.01	0.80 ± 0.01	-----
<i>B. subtilis</i>	100 ± 0.23	9.00 ± 0.03	0.64 ± 0.03
	50 ± 0.10	7.00 ± 0.01	81 ± 0.23
	25 ± 0.10	3.20 ± 0.01	49 ± 0.21
	12.50 ± 0.01	-----	10.24 ± 0.01
	6.25 ± 0.01	-----	-----
<i>C. albicans</i>	100 ± 0.23	7.00 ± 0.01	49 ± 0.10
	50 ± 0.10	4.30 ± 0.01	18.49 ± 0.10
	25 ± 0.10	3.50 ± 0.01	12.25 ± 0.01
	12.50 ± 0.01	2.50 ± 0.01	6.25 ± 0.01
	6.25 ± 0.01	-----	-----

The spectrum of activity of the standard antibiotic gentamycin against some specific bacteria is presented in Table 4. The IZD values of *S. aureus* ranged from 9.00 ± 0.01mm to 41.00 ± 0.20 mm, while *B. subtilis* ranged from 8.00 ± 0.01 mm to 35.00 ± 0.03 mm without any significant difference ($p < 0.05$).

Table 4. Spectrum of activity of the standard antibiotic Gentamycin against bacteria (Inhibition zone diameter IZD, mean ± SD)

Microorganism	Concentration of extract (mg/ml)	IZD = L (mm)	L ² (mm)
<i>S. aureus</i>	100.00 ± 0.23	41.00 ± 0.20	1681 ± 0.20
	50.00 ± 0.21	32.60 ± 0.10	1062 ± 0.21
	25.00 ± 0.10	25.60 ± 0.10	655 ± 0.20
	12.50 ± 0.01	20.00 ± 0.01	400 ± 0.10
	6.25 ± 0.01	9.00 ± 0.01	81 ± 0.10
<i>B. subtilis</i>	100.00 ± 0.23	35.00 ± 0.03	1225 ± 0.03
	50.00 ± 0.10	30.00 ± 0.01	900 ± 0.23
	25.00 ± 0.10	20.00 ± 0.01	400 ± 0.21
	12.50 ± 0.01	14.00 ± 0.10	196 ± 0.01
	6.25 ± 0.01	8.00 ± 0.01	-----

The activity of clotrimazole was recorded in Table 5. The IZD mm without any significant difference ($p < 0.05$). values recorded ranged from $6.00 \pm 0.01\text{mm}$ to 28.00 ± 0.02

Table 5. Activity of the standard antifungal agent Clotrimazole against the Fungi (IZDs)

Test organism	Concentration of clotrimazole (mg/ml)	IZD (mm)	IZD ² (mm)
<i>C. albicans</i>	100.00 ± 0.20	28.00 ± 0.02	784.00 ± 0.20
	50.00 ± 0.10	21.00 ± 0.01	441.00 ± 0.20
	25.00 ± 0.01	15.00 ± 0.01	225.00 ± 0.02
	12.50 ± 0.01	9.00 ± 0.02	81.00 ± 0.01
	6.25 ± 0.01	6.00 ± 0.01	36 ± 0.01

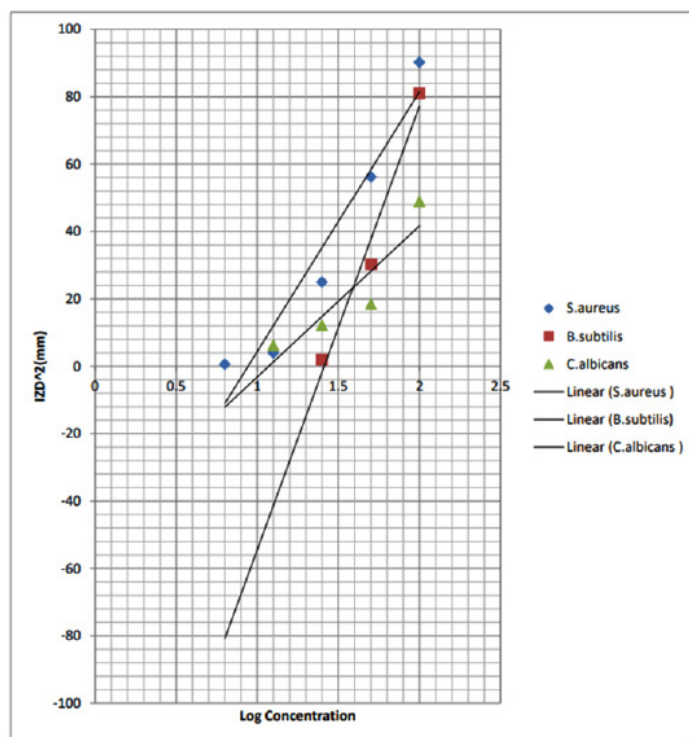


Figure 1. A graph of IZD² against concentrated methanolic extracts of *Bridelia ferruginea* for susceptible organisms.

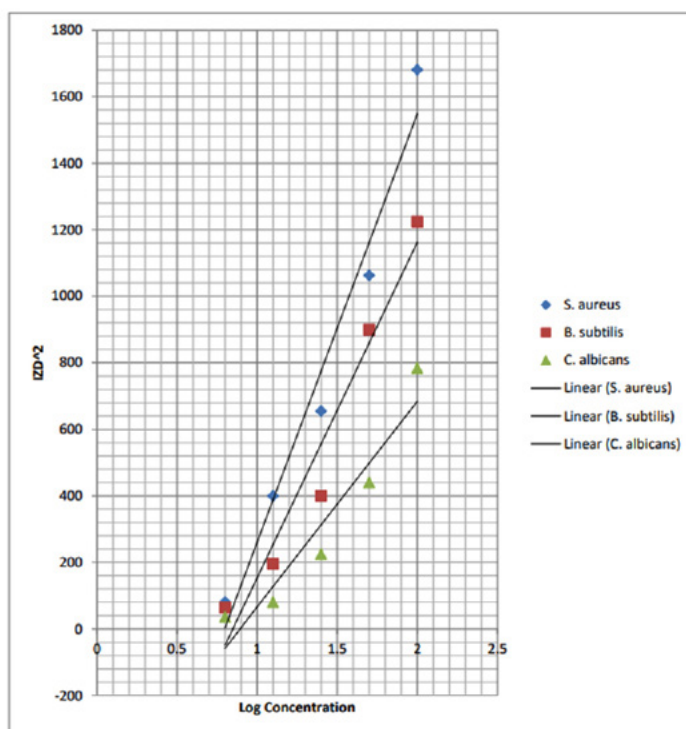


Figure 2. A graph of IZD² against log concentration of standard antibiotics (Gentamycin and clotrimazole).

The antimicrobial activity was more pronounced against bacteria than fungi. The MIC illustrates a decreasing inhibitory effect of the leaf extracts as the concentration decreases. This implies that antimicrobial activity of a substance is concentration-dependent and a function of the active ingredient reaching an organism [49]. From the results, the extract seems to be effective only against Gram positive bacteria and not Gram-negative bacteria. The reason for the different sensitivity between Gram positive and Gram-negative bacteria could be ascribed to the morphological differences between these microorganisms. Gram negative bacteria have an outer phospholipidic membrane carrying the structural

lipopolysaccharide components, which makes the cell wall partially impermeable. The Gram-positive bacteria are thus, more susceptible, having only an outer peptidoglycan layer which is not an effective permeability barrier. From the minimum inhibitory concentration, it revealed that it has greatest activity on *Staphylococcus aureus* (MIC = 8.91), followed by *Candida albicans* (MIC = 12.02mg/ml) and then, *B. subtilis* (MIC = 25.12) on which it has the least effect. On comparing the methanolic extract of the *Bridelia ferruginea* with the standard drugs (Gentamycin and Clotrimazole), it is clear that the standard drugs have greater activity [50].

Table 6. MIC determination from the graph for the standard antibiotics (Gentamycin and clotrimazole)

Test organism	Intercept on log conc axis	Antilog of intercept (MIC) mg/ml
<i>S. aureus</i>	0.80	6.31
<i>B. subtilis</i>	0.85	7.08
<i>C. albicans</i>	0.90	7.94

The result of Phytochemical analysis of the leaves of *Bridelia ferruginea* showed the presence of secondary metabolites which are: glycosides, flavonoids, alkaloids, saponins, tannins, proteins, carbohydrates, reducing sugars, steroids, and resins. From the results, it showed that alkaloids

and proteins were present in high concentration while glycosides, flavonoids, saponins, tannins, carbohydrate, and steroids were present in moderate concentration, whereas reducing sugar and resins were present in low concentration. The methanolic extract was found to possess antimicrobial

activity. It has effect on both Bacteria and fungi but not on Gram negative bacteria.

Previous studies demonstrated the presence of flavonoid components. Other authors showed the presence of terpenes and isomers of nepetalactone in *Nepeta cataria* and their bacteriostatic and fungistatic properties [51]. However, it is difficult to compare the data of this study with literature because several variables influence the results, such as the type of plant used, the climatic and environmental factors of the plant, the method of extraction and the use of appropriate solvents. The presence of these chemical constituents underscores the importance of these plants in

medicine. According to Prasathkumar et al, [52], all alkaloids contain quinine, morphine or reserpine which are used for malaria, pain relief and valuable tranquilizers respectively. Alkaloids are one of the largest groups of phytochemicals in plants, having amazing effects on humans and has led to the development of powerful pain killer medications. This suggests that *B. ferruginea* can be a useful source for malaria drug, pain relief and tranquilizer. The leaf extract has been reported to reduce plasma glucose levels in diabetic patients [53]. However, further experimental and research efforts on the plants and their extracts are needed to be able to specify the pharmacological implication.

Table 7. Phytochemical analysis

CONSTITUENTS	
1. General test for glycosides	++
2. Flavonoids	++
3. Alkaloids	+++
4. Saponins	++
5. Tannins	++
6. Resins	+
7. Proteins	+++
8. Carbohydrates	++
9. Reducing sugars	+
10. Fat and oils	-
11. Steroids	++
12. Terpenoids	-
13. Acidic compounds	Neutral
14. Starch	-

KEY:

- ABSENT

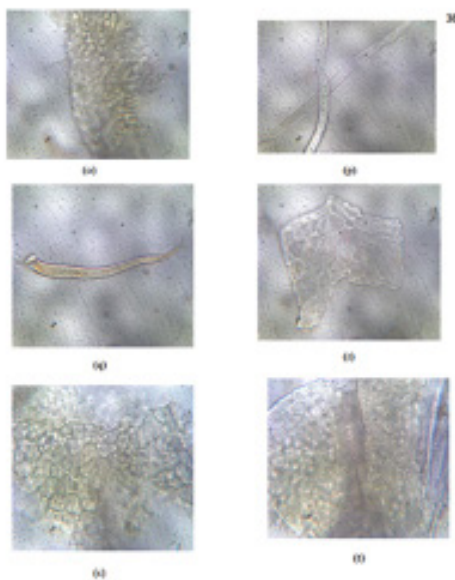
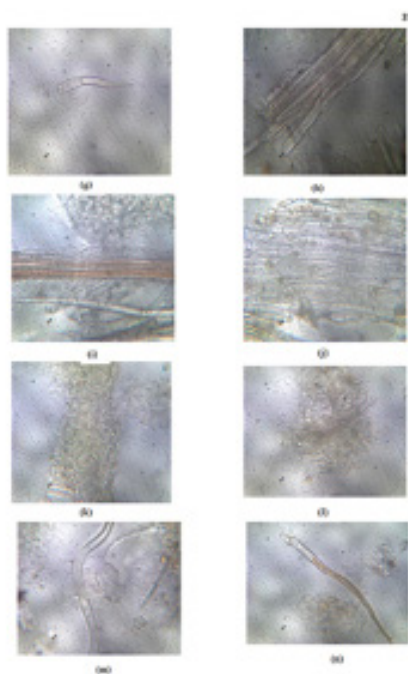
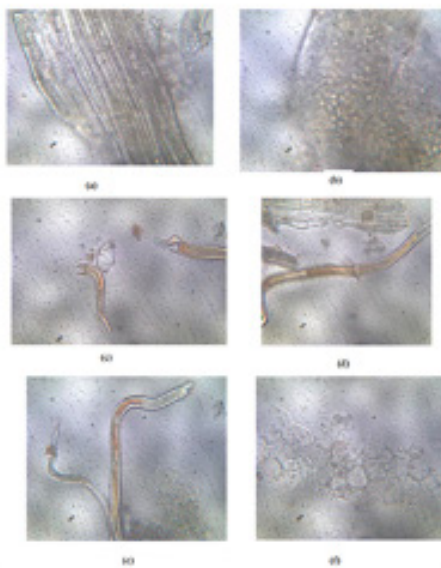
+ PRESENT

++ MODERATELY PRESENT

+++ ABUNDANTLY PRESENT

The microscopic evaluation showed the presence of prism calcium oxalate, bundle of fibers, circular palisade cells, trichomes, annular xylem vessels, epidermal cells with no stomata, large phloem fibers. The standardization of crude drugs has become very necessary and important because they are used in quality control especially to determine the

quality, purity, strength and if adulterated, the nature of the adulterants. It has also been used to determine the percentage yield of the crude *Bridelia ferruginea* material as indicated in the work and also in solving the problem of spoilage of herbal drugs.



- a) Bundle of annular xylem vessels and group of fibres
 b) Circular palisade cells
 c) Clothing unicellular trichome and a prism calcium oxalate
 d) Elongated clothing multicellular trichome
 e) Elongated non glandular trichome
 f) Elongated non glandular trichome
 g) Fractured multicellular trichome
 h) Group of fibres
 i) Large annular xylem vessels
 j) Large phloem fibres
 k) Large upper epidermal cells with no trichome
 l) Lower epidermis showing anomocytic type of stomata
 m) Multicellular elongated trichome
 n) Multicellular trichome
 o) Numerous palisade cells
 p) Pair of elongated unicellular non glandular trichome
 q) Unicellular clothing trichome
 r) Upper epidermis showing cicatrix
 s) Upper epidermis showing palisade cells with no stomata
 t) Upper epidermis with attached trichome

The result of the standardization test is presented in Table 8. The moisture content of the leave extract was 1.05 ± 0.02 , the total ash value was 5.00 ± 0.01 , the sulphated ash and acid insoluble ash ranged from 2.00 ± 0.01 to 1.15 ± 0.01 respectively.

Table 7. Standardization

Parameter	Result
Moisture content	1.05 ± 0.02
Total ash	5.00 ± 0.01
Acid insoluble ash	1.15 ± 0.01
Sulphated ash	2.00 ± 0.01

CONCLUSION

Pharmacognostic studies performed on the leaves support most of the claims with a few traditional uses yet to be approved. The study indicates the enormous potential of *B. ferruginea* with anti-microbial properties. Due to its wide application, the phytochemicals present has been documented, and they could be useful in lead compound discovery. A lot of untapped potentials for further investigation still remains despite its wide application.

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CONFLICT OF INTEREST

Authors declare no conflict of interest.

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