

Research Article

Influence of Plant Extracts on acceptability of Chilled Poultry Meat

Diaa Eldin IM¹, Eman T Abou Sayed-Ahmed¹, Hamada M Hassan¹, Fahim Shaltout^{2,*}, Gehan

Abdallah El-shorbagy¹

¹Department of Food Science, Faculty of Agriculture, Zagazig University, Egypt ²Department of Food Control, Faculty of Veterinary Medicine, Benha University, Egypt

ABSTRACT

Poultry meat is usually marketed at refrigerated temperatures (1 to $4 \circ$ C). Acceptability, Quality and Safety of refrigerated Poultry meat is the main concern for consumers and retailers. Poultry meat is contaminated from different sources as during slaughtering, during different manufacturing processes and during storage causing undesirable changes, microbial growth, spoilage and economic losses. Thus, the present study aimed to assess the effects of plant extracts (1.0 % Laurel, 1.0 % Moring and 1.0 % Olive leaf extract) on acceptability of raw Poultry breast meat stored at 1 to 4 ° C for 16.0 days. The Data revealed that samples treated with 1.0 % laurel, 1.0 % Moringa and 1.0 % Olive extracts maintained the acceptability until 16th, 1±th, 12thdays of chilling at 1 to 4 °C, respectively. Compared to untreated one which got spoiled by 6th day of chilling at 1 to 4 °C. Samples treated with plant extracts revealed significant decrease in their keeping quality tests and marked decrease in bacterial examination (total bacterial, total coliform, total Staphylococci) revealed that plant extracts have good antioxidant and antibacterial effects. Best effect was obtained in samples treated with 1.0 % laurel extract followed by samples treated with 1.0 %Moringa and those treated with 1.0 % olive leaf extract.

Keywords: acceptability, olive leaf extract, Poultry meat, Laurel, Moringa.

INTRODUCTION

Poultry meat has nutritional characters, as low fat content and high concentration of polyunsaturated fatty acids [1-5]. Poultry meat is low calorie food for its low fat content, its muscle lipids are highly subjected to oxidation due to the high unsaturation degree. Oxidation leads to deterioration in meat color, flavor, texture, nutrient losses, and poor shelf life. Simultaneously, some internal factors as iron content, antioxidant enzymes and external Factors as stress, temperature, feeding with highly oxidized feeds, slaughtering process, storage conditions, further processing steps, etc. which plays an important role in oxidation process of

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*Corresponding Author

Fahim Shaltout

Department of Food Control, Faculty of Veterinary Medicine, Benha University, Egypt, Tel: 00201006576059, ORCID: 0000-0002-8969-2677

E-mail: fahim.shaltout@fvtm.bu.edu.eg; fahimshaltout@hotmail.com

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Copyright: Diaa Eldin IM, et al. © (2023). This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited. poultry meat [3,6-10]. Poultry meat is a good protein sources, but after slaughtering, protein of these meat can be oxidized initiating a secondary lipid oxidation of products and causes loss of functional properties, acceptability and quality of meat protein [7,11].

Molecular oxygen undergoes a chain of reactions that leads to generation of free radicals. Under normal physiologic conditions a small percent of the oxygen that consumed during the metabolic reaction is changed to free radicals. These free radicals especially reactive nitrogen species and reactive oxygen species play a key regulatory role in homeostatic processes by interacting with fatty acids, proteins and nucleic acids. They act as transitional agents in essential oxidation– reduction reaction [12-15].

Primarily, when the production of reactive oxygen species and reactive nitrogen species doesn't exceed the capability of endogenous antioxidant barriers in the body, it implements beneficial functions and defense against invading pathogens. In contrast, when there is an excess and low activity of antioxidant defence, it potentially causes damage of the cellular components, induces destructive autoimmune responses and causes oxidative stress [16-19].

Free radical activities in meat is so important because high levels of reactive oxygen species might reduce meat sensory quality [20,21], reduction of essential amino acids like tryptophan and phenylalanine [22,23]. And loss of protein functions. As well as, degradation of polyunsaturated fatty acids section of meat lipids and diversion of oxymyoglobin [oxyMb (Fe2+)] to metmyoglobin [MetMb (Fe3+)] lead to generation of free radicals might result in deterioration of meat proteins [24,25]. To prevent protein and fat oxidations occurring in poultry meat is the use of antioxidants [26], consumers and producers tended to the natural alternatives from plant sources [27].

Due to their high phenolic content, spices, fruits, oilseeds, vegetables and grains seem to be good sources of natural antioxidants as a substitution to the synthetic ones [28]. The use of natural antioxidant extracts has been reported to raise meat tenderness [29]. Plant extracts to enhance the oxidative stability and acceptability of the poultry products [30]. Plants and their extracts have variable concentrations of phenolic compounds are thus regarded as an efficient source of antioxidants for controlling oxidation reactions [31,32].

Laurusnobilis is an evergreen plant with 2–3 m high and a pair of stems. Alpha-tocopherol is the foremost isomer in its vegetative parts. Leaves comprise flavonoids, phenolic acids and isoquinoline alkaloids. Moreover, leaves and roots are rich in alpha-tocopherol and flavonoids [33,34]. Antimicrobial activity of laurel leaf extract has importance as a natural antioxidant as they are part of human diet and its biodegradability leads to low poisonous residue problems [35,36]. L. nobilis has powerful antimicrobial activity against 20 strains of bacteria as Staphylococcus aureus, Escherichia coli, Listeria monocytogenes and Salmonella [37,38].

Moringaoleifera leaves have high quantities of ascorbic acid, phenolic, carotenoids and flavonoids [39, 40]. So, it is considered as potent antimicrobial and antioxidant. M. oleifera leaf extract keep goat meat patties from the oxidative rancidity [35,41]. As well as it significantly extend ghee shelf life [39,42]. The leaves having antifungal and antimicrobial characteristics, therefore, have a proven history in food applications as a biopreservative and neutraceuticals [43].

Olive leaves are observed as predominantly rich sources of phenolic composites [44,45]. Their chief biological active compounds are classified into oleuropeosides, simple phenolics and flavonoids [46,47]. Phenolic compounds can prevent the growth and secretion of Staphylococcus aureus enterotoxins [8,9,48-51].

In this study, application of natural bioactive compounds from natural plants to inhibit lipid and protein oxidation, improve oxidative stability and acceptability of Poultry meat.

MATERIALS AND METHODS

Preparation of Laurel Leaf Extract (1.0 % LLE) [52].

Preparation of Moringa Leaf Extract (1.0 % MLE) [53].

Preparation of Olive Leaf Extract (1.0 % OLE)

According to the method recommended by [54].

Preparation of samples

Poultry breast meat slices (100.0 grams), two cms in thickness. Samples were placed in a separate sterile plastic bags in an icebox and carried out to the Laboratory without undue delay under complete aseptic conditions. Poultry meat samples were divided into two main groups (treated and untreated). Treated groups were subdivided into three groups (27 samples for each). First group samples were dipped in 1.0 % Laurel Leaf Extract for five minutes with proper mixing,

2ndgroup samples were dipped in 1.0 % Moringa Leaf Extract for five minutes and 3rdgroup samples in 1.0 % Olive Leaf Extract. All samples were stored chilled at 1 to 4 \circ C and examined every 48 hours (0. 2nd ,4th, 6th, 8th, 10th,12th, 14th, 16th) for acceptability. The experiment was replicated three times/day and the examination repeated every 48 hours for 16 days of chilling at 1 to 4 °C.

SENSORY EXAMINATION AND ACCEPTABILITY

Bacteriological investigation [19,48,49].

Chemical Examination

PH detection

Total Volatile Nitrogen (TVN) (mg/100 g)

Thiobarbituric Acid "TBA" examination

Statistical Analysis

The Data analysis by using SPSS statistical software program (SPSS for Windows version 16, Spss Inc., USA).

RESULTS

Sensory and Acceptability examination:

Poultry meat samples stored at chilling 1 to 4 °C revealed that untreated samples were completely spoiled by 6th day of chilling at 1 to 4 °C. Addition of 1.0 % Laurel Leaf Extract maintained the whole acceptability of sensory parameters until 16th day, 1.0 % Moringa Leaf Extract maintained the acceptability until 14th day while1.0 % Olive Leaf Extract conserved the acceptance of sensory parameters till 12thday of chilling at 1 to 4 °C (Table A).

Table A. Acceptance (color, odor and texture) of fresh Poultry breast meat samples treated with plant extracts during chilling at 1 to 4 • C (mean ± standard error "SE").

Groups -	Acceptability of Poultry breast meat during chilling at 1 to 4 $^\circ C$										
	Zero day	2 nd day	4 th day	6 th day	8 th day	10 th day	12 th day	14 th day	16 th day		
Untreated	9.000 ± 0.00 ^{Aa}	6.330 ± 0.33 ^{Bc}	5.670 ± 0.33 ^{Bb}	4.000 ± 0.58 ^{Cc}	2.000 ± 0.00 ^{Dc}	1.000± 0.00 ^{Dc}					
1.0 % LLE	9.000 ± 0.00 ^{Aa}	9.000 ± 0.00^{Aa}	8.000 ± 0.00^{Ba}	7.330 ± 0.33^{Ca}	6.330 ± 0.33 ^{Da}	6.330 ± 0.33^{Da}	6.000 ± 0.00^{Da}	5.000 ± 0.00^{Ea}	4.33± 0.33 ^{Fa}		
1.0 % MLE	9.00 ± 0.00^{Aa}	$8.00\pm0.00^{\text{Bb}}$	8.00 ± 0.00^{Ba}	7.00 ± 0.00^{Cab}	7.00 ± 0.00^{Ca}	6.00 ± 0.00^{Da}	5.00 ± 0.00^{Eb}	4.33 ± 0.33^{F_a}			
1.0 % OLE	9.000 ± 0.00 ^{Aa}	8.330 ± 0.33 ^{Aab}	7.330 ± 0.33 ^{Ba}	6.000 ± 0.00 ^{Cb}	5.330 ± 0.33 ^{Cb}	4.330± 0.33 ^{Db}	$4.000 \pm 0.00^{\text{Dc}}$				

Score system:

9: Excellent. 8: Very very good. 7: Very good. 6: Good. 5: Medium.

4: Fair. 3: Poor 2: Very poor. 1: Very very poor

Mean values with different superscript Capital letters in the same row are significantly different at (P<0.05). Mean values with different superscript Small letters in the same column are significantly different at (P<0.05).

Bacteriological investigation of Poultry breast meat samples:

Untreated group revealed highest Aerobic Bacterial Count (TBC), Coliform Count (TCC) and Staphylococci Count (TSC). Treated samples revealed significant gradual decrease in all these counts during chilling at 1 to 4°C. Counts were the

lowest in samples treated with 1.0 % laurel extract followed by those treated with 1.0 % moringa extract and finally samples treated with 1.0 % olive extract. Our Data revealed that 1.0 % Laurel Leaf Extract, 1.0 % Moringa Leaf Extract and 1.0 % Olive Leaf Extract have a positive impact in decreasing all previously mentioned counts in treated samples compared to untreated one indicating their potent antibacterial effect (Tables B,C, &D).

Samples treated with 1.0 % Laurel extracts revealed the highest reduction percentage of TBC 99.980 % and 99.990% at 6th and 8th day, respectively. Samples treated with 1.0 % Olive Leaf Extract revealed the highest reduction % of TCC 99.870 % and 99.990 % at 6thand 8thday, respectively. Smples treated with laurel extract revealed the highest reduction percent of TSC 99.52% and 99.930 % at 6thand 8thday, respectively.

Table B. Pattern of Aerobic Plate Count (log10 cfu/g) in Poultry breast meat samples treated with plant extracts during chilling at 1 to 4 ∘ C (mean ± standard error "SE").

	Acceptability of Poultry breast meat during chilling at 1 to 4 $^\circ\text{C}$											
Zero day	2 nd day	4 th day	6 th day	8 th day	10 th day	12 th day	14 th day	16 th day				
9.000 ± 0.00 ^{Aa}	6.330 ± 0.33^{Bc}	5.670 ± 0.33^{Bb}	4.000 ± 0.58^{Cc}	2.000 ± 0.00 ^{Dc}	1.000 ± 0.00^{Dc}							
9.000 ± 0.00 ^{Aa}	9.000 ± 0.00^{Aa}	8.000 ± 0.00^{Ba}	7.330 ± 0.33^{Ca}	6.330 ± 0.33^{Da}	6.330 ± 0.33^{Da}	6.000 ± 0.00^{Da}	5.000 ± 0.00^{Ea}	4.33± 0.33 ^{Fa}				
9.00± 0.00 ^{Aa}	8.00± 0.00 ^{Bb}	8.00 ± 0.00^{Ba}	7.00± 0.00 ^{Cab}	7.00 ± 0.00^{Ca}	6.00 ± 0.00^{Da}	5.00± 0.00 ^{Eb}	4.33 ± 0.33^{Fa}					
9.000 ± 0.00 ^{Aa}	8.330 ± 0.33^{Aab}	7.330 ± 0.33^{Ba}	$6.000 \pm 0.00^{\text{Cb}}$	5.330 ± 0.33 ^{Cb}	4.330± 0.33 ^{Db}	4.000± 0.00 ^{Dc}						

Mean values with different superscript Capital letters in the same row are significantly different at (P<0.05). Mean values with different superscript Small letters in the same column are significantly different at (P<0.05).

Table C. Pattern of Coliform Count (log10 cfu/g) in Poultry breast meat samples treated with plant extracts during chilling at 1 to 4 ∘ C (mean ± standard error "SE").

6	Total Coliform Count (log10 cfu/g) in Poultry breast meat samples treated with plant extracts during chilling at 1 to 4 \circ C										
Groups	Zero day	2 nd day	4 th day	6 th day	8 th day	10 th day	12 th day	14 th day	16 th day		
untreated	4.80± 3.46 ^{Ca}	5.14± 4.02 ^{BCa}	5.29 ± 4.28^{BCa}	5.53 ± 4.20^{Ba}	5.57 ± 5.28^{Aa}						
1.0 % LLE	4.57 ± 3.28^{Ac}	4.30± 3.28 ^{Cd}	3.60± 3.11 ^{Eb}	2.75± 2.14 ^{EFb}	2.45± 2.08 ^{Fb}	2.25± 2.02 ^{Fb}	2.15 ± 2.02^{Fc}	4.20± 3.31 ^{Db}	4.43 ± 3.57^{Ba}		
1.0 % MLE	4.68± 3.14 Ab	4.47 ± 3.14^{Bc}	4.13± 3.11 ^{cb}	2.80± 2.57 ^{Db}	2.65± 2.08 ^{Db}	2.35± 2.02 ^{Db}	4.19± 3.14 ^{Cb}	4.50 ± 3.57^{Ba}			
1.0 % OLE	4.72± 3.28 ^{Aab}	4.57± 3.43 ^{Bb}	4.23± 3.28 ^{bb}	3.7± 2.11 ^{Eb}	2.50± 2.02 ^{Eb}	3.19 ± 3.14^{Da}	4.40± 3.23 ^{ca}				

Mean values with different superscript Capital letters in the same row are significantly different at (P<0.05). Mean values with different superscript Small letters in the same column are significantly different at (P<0.05).

Table D. Pattern of Staphylococci Count (log10 cfu/g) in Poultry meat samples treated with plant extractsduring chilling at 1 to 4 • C (mean ± standard error "SE")

Groups	Total Staphylococci Count (log10 cfu/g) in Poultry meat samples treated with 3 different plant extracts during chilling at 1 to 4 $^\circ$ C										
	Zero day	2 nd day	4 th day	6 th day	8 th day	10 th day	12 th day	14 th day	16 th day		
Untreated	5.23± 4.17 ^{Ba}	5.39± 4.25 ^{Ba}	5.59 ± 4.23^{Ba}	6.13 ± 5.14^{Ba}	6.50 ± 5.11^{Aa}						
1.0 % LLE	5.11 ± 4.08^{Ac}	4.26± 3.23 ^{Cb}	4.14± 2.57 ^{DEb}	3.65± 2.86 ^{EFb}	3.35± 2.86 ^{Fb}	2.80± 2.57 ^{Fb}	2.25± 2.02 ^{Fc}	4.22± 3.11 ^{CDb}	5.42± 3.88 ^{Ba}		
1.0 % MLE	5.13± 4.17 ^{Ac}	4.28± 3.25 ^{cb}	4.14± 3.17 ^{CDb}	3.65± 2.86 ^{CDb}	3.50± 2.57 ^{CDb}	2.35 ± 2.08^{Eb}	4.13± 3.20 ^{CDb}	4.51 ± 4.65^{Ba}			
1.0 % OLE	5.18± 4.8 ^{Ab}	4.28± 3.17 ^{Cb}	4.18± 3.17 ^{CDb}	$3.70 \pm 3.11^{\text{Db}}$	3.66± 3.27 ^{Db}	$4.18 \pm 3.20^{\text{CDa}}$	4.53 ± 3.14^{Ba}				

Mean values with different superscript Capital letters in the same row are significantly different at (P<0.05). Mean values with different superscript Small letters in the same column are significantly different at (P<0.05).

Chemical Examination

Differences in pH mean value between untreated and treated samples are significant (P<0.05) (Table D) untreated group revealed the highest pH 6.880 ± 0.01 at 6th day of chilling at 1

to 4 °C compared to treated groups which revealed lower pH., samples treated with laurel extract revealed the lowest pH value 5.920 ± 0.01 followed by samples treated with Moringa Leaf Extract 6.010 ± 0.020 and finally Olive Leaf Extract 6.130 ± 0.020 . PH decreased may be owing to the antioxidant effect of plant extracts as mentioned before pH is one of the factors that is associated with lipid oxidation in meat.

Used plant extracts resulted in decrease of TVN values in

Poultry meat samples with significant differences when compared to untreated group that means it decreased protein oxidation in Poultry breast meat. Highest TVN values were recorded in untreated group 19.870 ± 0.050 at 6th day of chilling at 1 to 4 °C while lowest one was recorded in sample treated with 1.0 % laurel extract 10.130 ± 0.050 followed by 1.0 % Moringa Leaf Extract 11.520 ± 0.080 then, 1.0 % Olive Leaf Extract 14.490 ± 0.050 . (Table E).

By the same way, an increase in TBA value was observed in untreated group and lowered TBA values in treated groups with a highly significant difference when compared to untreated one. Highest TBA value was recorded in untreated group 0.86 ± 0.01 at 6th day of chilling at 1 to 4 °C compared to $0.330 \pm 0.010, 0.390 \pm 0.010$ and 0.500 ± 0.010 which recorded in samples treated by 1.0 % laurel, 1.0 % moringa and 1.0 % olive extract, respectively (Table F & G).

Table E. Effect of plant extracts on PH value applied in Poultry breast meat samples during chilling at 1 to 4 °C(mean ± standard error "SE").

	PH values of Poultry breast meat samples during chilling at 1 to 4 $^\circ C$									
Groups	Zero day	2 nd day	4 th day	6 th day	8 th day	10 th day	12 th day	14 th day	16 th day	
Untreated	5.700 ± 0.01^{Ea}	6.100 ± 0.02^{Da}	6.180 ± 0.02^{Ca}	6.880 ± 0.01^{Ba}						
1.0 % LLE	5.630 ± 0.00^{1c}	$5.700 \pm 0.01^{\text{Hd}}$	5.800 ± 0.02^{Gd}	5.920 ± 0.01^{Fa}	6.010 ± 0.02^{Ea}	$6.140 \pm 0.02^{\text{Dc}}$	6.260 ± 0.02^{Cc}	$6.380 \pm 0.03^{\text{Bb}}$	6.610 ± 0.04^{Aa}	
1.0 % MLE	$5.650 \pm 0.01^{\text{Hb}}$	5.740 ± 0.01^{Gc}	5.880 ± 0.01^{Fc}	$6.010 \pm 0.02^{\text{Ec}}$	$6.140 \pm 0.03^{\text{Dc}}$	6.320 ± 0.02 ^{Cb}	$6.410 \pm 0.01^{\text{Bb}}$	$6.510 \pm 0.02^{\text{Aa}}$		
1.0 % OLE	5.660 ± 0.01^{Gb}	$5.780 \pm 0.01^{\text{Fb}}$	$5.960 \pm 0.01^{\text{Eb}}$	$6.130 \pm 0.02^{\text{Db}}$	6.290 ± 0.01 ^{Cb}	$6.480 \pm 0.02^{\text{Ba}}$	6.690 ± 0.02^{Aa}			

Mean values with different superscript Capital letters in the same row are significantly different at (P<0.05). Mean values

with different superscript Small letters in the same column are significantly different at (P<0.05)

Table F. Effect of plant extracts on TVN value (mg/100 g) applied in Poultry breast meat samples during chilling at 1 to 4 °C(mean ± standard error "SE").

Groups –	TVN values of Poultry breast meat samples during chilling at 1 to 4° C										
	Zero day	2 nd day	4 th day	6 th day	8 th day	10 th day	12 th day	14 th day	16 th day		
Untreated	3.120 ± 0.03 ^{Ea}	8.860 ± 0.12 ^{Da}	14.940 ± 0.07^{Ca}	$19.870 \pm 0.05^{\text{Ba}}$							
1.0 % LLE	2.910± 0.02 ^{Hb}	5.210± 0.03 ^{Gd}	8.470 ± 0.05^{Fd}	10.130± 0.05 ^{Ed}	$12.970 \pm 0.12^{\text{Dd}}$	17.480± 0.19 ^{cc}	19.660± 0.10 ^{Bc}	21.680 ± 0.18^{Ab}	21.910 ± 0.23 ^{Aa}		
1.0 % MLE	2.950 ± 0.02 ^{Gb}	5.490 ± 0.03 ^{Fc}	9.250 ± 0.07^{Ec}	11.520 ± 0.08 ^{Dc}	14.580 ± 0.14 ^{Cc}	19.840 ± 0.06 ^{вь}	22.250 ± 0.19 ^{Ab}	22.360 ± 0.15 ^{Aa}			
1.0 % OLE	3.070 ± 0.02 ^{Fa}	6.060 ± 0.02 ^{Eb}	11.820 ± 0.07 ^{Db}	14.490 ± 0.05 ^{cb}	18.510 ± 0.13 ^{Bb}	22.850 ± 0.09 ^{Aa}	23.070 ± 0.13 ^{Aa}				

Mean values with different superscript Capital letters in the same row are significantly different at (P<0.05). Mean values

with different superscript Small letters in the same column are significantly different at (P<0.05)

Table G. Effect of plant extracts on TBA (mg of malondialdehyde / kg)applied in Poultry breast meatsamples during chilling at 1 to 4 °C(mean ± standard error "SE").

Groups –	TVN values of Poultry breast meat samples during chilling at 1 to 4° C										
	Zero day	2 nd day	4 th day	6 th day	8 th day	10 th day	12 th day	14 th day	16 th day		
- Untreated	3.120 ± 0.03 ^{Ea}	8.860 ± 0.12 ^{Da}	14.940 ± 0.07 ^{Ca}	19.870 ± 0.05 ^{Ba}							
1.0 % LLE	2.910± 0.02 ^{Hb}	5.210± 0.03 ^{Gd}	8.470± 0.05 ^{Fd}	10.130± 0.05 ^{Ed}	$12.970 \pm 0.12^{\text{Dd}}$	17.480± 0.19 ^{Cc}	19.660± 0.10 ^{Bc}	$21.680 \pm 0.18^{\rm Ab}$	21.910 ± 0.23 ^{Aa}		
1.0 % MLE	2.950 ± 0.02 ^{Gb}	5.490 ± 0.03 ^{Fc}	9.250 ± 0.07^{Ec}	11.520 ± 0.08 ^{Dc}	14.580 ± 0.14 ^{Cc}	19.840 ± 0.06 ^{Bb}	22.250 ± 0.19 ^{Ab}	22.360 ± 0.15^{Aa}			
1.0 % OLE	3.070 ± 0.02 ^{Fa}	6.060 ± 0.02 ^{Eb}	11.820 ± 0.07 ^{Db}	14.490 ± 0.05 ^{cb}	18.510 ± 0.13 ^{Bb}	22.850 ± 0.09 ^{Aa}	23.070 ± 0.13 ^{Aa}				

Mean values with different superscript Capital letters in the same row are significantly different at (P<0.05). Mean values

with different superscript Small letters in the same column are significantly different at (P<0.05)

DISCUSSION

Sensory evaluation is quick, efficient and easy method for getting an idea about acceptance and quality of the product. it depends on organoleptic characteristics as color, odor, texture and product acceptability [55-57]. Table (A) revealed that sensory evaluation of treated samples was improved and extended shelf-life during chilling at 1 to 4 °C. The obtained Data revealed that the best sensory quality was achieved in Poultry samples treated with 1.0 % laurel extract, good enhancement of sensory quality in Poultry samples treated with 1.0 % moringa extract followed by those treated with 1.0 % olive extract as compared to untreated samples and these Data are similar to those results mentioned by [13,17,54,58]. According to their results laurel leaf extract used to extend shelf life of meat with improving the sensory parameters without causing undesirable odor or taste. The Data came in agreement with those mentioned by [41] which suggested addition of moringa extract to extend the shelf life of meat without any alteration in sensory quality of meat and those results of [50,59-61] their results revealed that 1.0 % olive leaf extract can maintain sensory parameters of meat when applied in poultry meat samples and retard microbial growth due to its antimicrobial effect.

The Data revealed that 1.0 % laurel extract has a potential applicability as a natural substitute to synthetic food preservatives to improve food safety and extend its shelf life confirming results in current study which agreed with those obtained by [62-64,69] they suggested application of natural plant extracts like moringa extract in meat and meat products contributing to its polyphenols content which are beneficial in antioxidant and antimicrobial functions.

Da Silveira SM, et al. [65] demonstrated that laurel extract applied in fresh sausages at concentration of 0.10 g/100.0 g can provide additional protection of the product against microbial growth, thus increasing its shelf life. The extract causes an obvious decline in pH, TVN and TBA having a potent antioxidant effect and these results are the same Data obtained in our study which came in the same line with [66] they revealed that pH, TVN and TBA values of Poultry patties were significantly decreased in samples treated with 100.0 grams of moringa leaf powder / kg. The Data are constant with those of [67,68] they conveyed that olive leaf extract can be used for meat preservation due to its antimicrobial and antioxidant effects thanks to its phenolic content.

Marked reduction in PH. TVN and TBA in poultry samples with Olive Leaf Extract indicating that Olive Leaf Extract is a powerful source of polyphenols having both antioxidant and antibacterial properties capable of decreasing microbial growth and increasing meat shelf-life similar to results revealed by[36,50,59-61,69].

CONCLUSION

Extracts of 1.0 % laurel, 1.0 % moringa and 1.0 % olive leaves maintained the sensory attributes of Poultry breast meat samples during chilling at 1 to 4 °C, possess considerable amounts of phenolic compounds exhibiting potent antimicrobial and antioxidant properties extend meat shelf life. So, addition of these plant extracts to meat and its products could improve the quality and acceptability of consumers.

REFERENCES

- Al Shorman AAM, Shaltout FA, Hilat N (1999):Detection of certain hormone residues in meat marketed in Jordan. Jordan University of Science and Technology, 1st International Conference on Sheep and goat Diseases and Productivity. pp. 23-25.
- Edris AM, Shaltout FA, Abd Allah AM. (2005). Incidence of Bacillus cereus in some meat products and the effect of cooking on its survival. Zag Vet J. 33(2):118-124.
- Edris AM, Hemmat MI, Shaltout FA, Elshater MA, Eman FMI. (2012). Study on incipient spoilage of chilled chicken cuts-up. Benha Vet Med J. 23(1):81-86.
- Nkukwana TT, Muchenje V, Masika PJ, Hoffman LC, Dzama K, Descalzo AM. (2014). Fatty acid composition and oxidative stability of breast meat from broiler chickens supplemented with Moringa oleifera leaf meal over a period of refrigeration. Food Chem. 142:255-261.
- Sobhy A, Shaltout F. (2020). Prevalence of some food poisoning bacteria in semi cooked chicken meat products at Qaliubiya governorate by recent Vitek 2 compact and PCR techniques. Benha Vet Med J. 38(2):88-92.
- Edris AM, Hemmat MI, Shaltout FA, Elshater MA, Eman FMI. (2012). Chemical analysis of chicken meat with relation to its quality. Benha Vet Med J. 23(1):87-92.
- Estévez M. (2015). Oxidative damage to poultry: from farm to fork. Poult Sci. 94(6):1368-1378.

- Shaltout FA, Lamada HM, Edris EAM. (2020). Bacteriological examination of some ready to eat meat and chicken meals. Biomed J Sci & Tech Res. 27(1):20461-20465.
- Shaltout FA, Mohammed IZ, Afify ESA. (2020). Bacteriological profile of some raw chicken meat cuts in Ismailia city, Egypt. Benha Vet Med J. 39(1):11-15.
- Sobhy A, Fahim S. (2020). Detection of food poisoning bacteria in some semi-cooked chicken meat products marketed at Qaliubiya governorate. Benha Vet Med J. 38(2):93-96.
- 11. Edris AM, Shaltout FA, Arab WS. (2005). Bacterial Evaluation of Quail Meat. Benha Vet Med J. 16(1):1-14.
- Gaschler R, Schwager S, Umbach V, Frensch P, Schubert T. (2014). Expectation mismatch: differences between selfgenerated and cue-induced expectations. Neuroscience & Biobehavioral Reviews. 46(Part1):139-157.
- Shaltout FA, El-diasty EM, Elmesalamy M, Elshaer M. (2014). Study on fungal contamination of some chicken meat products with special reference to 2 the use of PCR for its identification. Conference, Veterinary Medical Journal. 60:1-10.
- Shaltout FA, Salem RM, El-Diasty EM, Hassan WIM. (2019). Effect of Lemon Fruits and Turmeric Extracts on Fungal Pathogens in Refrigerated Chicken Fillet Meat. Global Veterinaria. 21(3):156-160.
- Shaltout FA, Nasief MZ, Lotfy LM, Gamil BT. (2019). Microbiological status of chicken cuts and its products. Benha Vet Med J. 37(1):57-63.
- Barbieri E, Sestili P. (2012). Reactive oxygen species in skeletal muscle signaling. J Signal Transduct. 2012:982794.
- Shaltout FA, El-diasty EM, Mohamed MS. (2014). Incidence of lipolytic and proteolytic fungi in some chicken meat products and their public health significance. 1st Scientific conference of food safety and Technology. pp. 79-89.
- Shaltout FA, El-diasty EM, Salem RM, Hassan MA. (2016). Mycological quality of chicken carcasses and extending shelf -life by using preservatives at refrigerated storage. Veterinary Medical Journal–Giza. 62(3):1-10.

- Shaltout FA, Thabet MG, Koura HA. (2018). Impact of Some Essential Oils on the Quality Aspect and Shelf Life of Meat Benha Vet Med J. 33(2):351-364.
- Bartosz G, Kołakowska A. (2010). Lipid oxidation in food systems. Chemical, biological, and functional aspects of food lipids. In: Sikorski Z, Kołakowska A. eds. CRC Press, New York, USA. pp. 163-184.
- Hassanin FS, Shaltout FA, Lamada HM, Abd Allah EM. (2011). The effect of preservative (nisin) on the survival of listeria monocytogenes. Benha Vet Med J. 2011(Special Issue [I]):141-145.
- 22. Ganhão R, Morcuende D, Estévez M. (2010). Tryptophan depletion and formation of α -aminoadipic and γ -glutamic semialdehydes in porcine burger patties with added phenolic-rich fruit extracts. J Agric Food Chem. 58(6):3541-3548.
- Saif M, Saad SM, Hassanin FS, Shaltout FA, Zaghlou M. (2019). Prevalence of methicillin-resistant Staphylococcus aureus in some ready-to-eat meat products. Benha Vet Med J. 37(1):12-15.
- 24. Hassan MA, Shaltout FA. (2004). Comparative Study on Storage Stability of Beef, Chicken meat, and Fish at Chilling Temperature. Alex J Vet Science. 20(21):21-30.
- Suman SP, Joseph P. (2013). Myoglobin chemistry and meat color. Annu Rev Food Sci Technol. 4:79-99.
- Descalzo AM, Sancho AM. (2008). A review of natural antioxidants and their effects on oxidative status, odor and quality of fresh beef produced in Argentina. Meat Sci. 79(3):423-436.
- Aziz SB, Abdulwahid RT, Rasheed MA, Abdullah OG, Ahmed HM. (2017). Polymer Blending as a Novel Approach for Tuning the SPR Peaks of Silver Nanoparticles. Polymers (Basel). 9(10):486.
- Shah MA, Bosco SJ, Mir SA. (2014). Plant extracts as natural antioxidants in meat and meat products. Meat Sci. 98(1):21-33.
- Chulayo A, Muchenje V. (2013). Effect of pre-slaughter conditions on physico-chemical characteristics of mutton from three sheep breeds slaughtered at a smallholder rural abattoir. South African Journal of Animal Science. 43(Suppl 2):S64-S68.

- Shahidi F, Zhong Y. 2010. Novel antioxidants in food quality preservation and health promotion. European Journal of Lipid Science and Technology. 112(9):930-940.
- Shaltout FA. (2002). Microbiological Aspects of Semicooked Chicken Meat Products. Benha Vet Med J. 13(2):15-26.
- Pennington JA, Fisher RA. (2009). Classification of fruits and vegetables. Journal of Food Composition and Analysis. 22(Suppl):S23-S31.
- Shaltout FA, Abdel Aziz AM. (2004). Salmonella enterica Serovar Enteritidis in Poultry Meat and their Epidemiology. Vet Med J-Giza. 52(3):429-436.
- 34. Farhoosh R, Tavakoli J, Khodaparast MHH. (2008). Chemical composition and oxidative stability of kernel oils from two current subspecies of Pistacia atlantica in Iran. Journal of the American Oil Chemists' Society. 85(8):723.
- Shaltout FA. (1998). Proteolytic Psychrotrophes in Some Meat products Alex. Vet Med J. 14(2):97-107.
- 36. Hamdan MS, Masoud WM. (2020). The antibacterial activity of Laurus nobilis leaf extract and its potential use as a preservative for fresh lamb meat. African Journal of Microbiology Research. 14(11):617-624.
- Shaltout FA. (2009). Microbiological quality of chicken carcasses at modern Poultry plant. The 3rd Scientific Conference, Faculty of Vet Med Benha University, Egypt. p. 1-3.
- Ouibrahim A, Tlili-Ait-Kaki Y, Bennadja S, Amrouni S, Djahoudi A, Djebar M. (2013). Evaluation of antibacterial activity of Laurus nobilis L., Rosmarinus officinalis L. and Ocimum basilicum L. from Northeast of Algeria. AJMR. 7(42):4968-4973.
- Anwar F, Latif S, Ashraf M, Gilani AH. (2007). Moringa oleifera: a food plant with multiple medicinal uses. Phytotherapy Research: PTR. 21(1):17-25.
- Hassanin FS, Hassan MA, Shaltout FA, Elrais-Amina M. (2014). Clostridium Perfringens in Vacuum Packaged Meat Products. Benha Vet Med J. 26(1):49-53.
- Jayawardana BC, Liyanage R, Lalantha N, Iddamalgoda S, Weththasinghe P. (2015). Antioxidant and antimicrobial activity of drumstick (Moringa oleifera) leaves in herbal

chicken sausages. LWT-Food Science and Technology. 64(2):1204-1208.

- Hassanin FS, Shaltout FA, Homouda SN, Arakeeb SM. (2019). Natural preservatives in raw chicken meat. Benha Vet Med J. 37(1):41-45.
- 43. Das AK, Rajkumar V, Verma AK, Swarup D. (2012). Moringa oleiferia leaves extract: a natural antioxidant for retarding lipid peroxidation in cooked goat meat patties. IFST. 47(3):585-591.
- 44. Silva S, Gomes L, Leitão F, Coelho AV, Boas LV. (2006). Phenolic Compounds and Antioxidant Activity of Olea europaea L. Fruits and Leaves. Food Science and Technology International. 12(5):385-395.
- 45. Shaltout FA, Elshater M, Abd El-Aziz WA. (2015). Bacteriological assessment of street vended meat products sandwiches in Kalyobia Governorate. Benha Vet Med J. 28(2):58-66.
- 46. Shaltout FA, Hasan MA, Hassanin FS. (2004). Thermal inactivation of Enterohaemorrhagic E. coli O157:H7 and its sensitivity to nisin and Lactic acid bacteria. The First Scientific Conference, Faculty of Vet Med, Moshtohor, Zagazig University Benha branch, Egypt.
- Botsoglou E, Govaris A, Ambrosiadis I, Fletouris D, Botsoglou N. (2014). Effect of olive leaf (Olea europea L.) extracts on protein and lipid oxidation of long-term frozen n-3 fatty acids-enriched pork patties. Meat Sci. 98(2):150-157.
- Shaltout FA, Zakaria IM, Nabil ME. (2018). Incidence of Some Anaerobic Bacteria Isolated from Chicken Meat Products with Special Reference to Clostridium perfringens. Benha Vet Med J. 33(2):292-304.
- Shaltout FA, Maarouf AAA, Ahmed EMK. (2018). Heavy Metal Residues in chicken cuts up and processed chicken meat products. Benha Vet Med J. 34(1):473-483.
- Saleh E, Morshdy AE, El-Manakhly E, Al-Rashed S, F Hetta H, Jeandet P, et al. (2020). Effects of Olive Leaf Extracts as Natural Preservative on Retailed Poultry Meat Quality. Foods. 9(8):1017.
- Afify SA, Shaltout F, Mohammed IZ. (2020). Detection of E. coli O157 and Salmonella species in some raw chicken meat cuts in Ismailia province, Egypt. Benha Vet Med J. 39(1):101-104.

- Dub AM, Dugani AM. (2013). Antithrombotic effect of repeated doses of the ethanolic extract of local olive (Olea europaea L.) leaves in rabbits. Libyan J Med. 8(1):20947.
- 53. Falowo A, Muchenje V, Hugo A, Aiyegoro O, Fayemi P. (2017). Antioxidant activities of Moringa oleifera L. and Bidens pilosa L. leaf extracts and their effects on oxidative stability of ground raw beef during refrigeration storage. CyTA-Journal of Food. 15(2):249-256.
- 54. Tometri SS, Ahmady M, Ariaii P, Soltani MS. (2020). Extraction and encapsulation of Laurus nobilis leaf extract with nano-liposome and its effect on oxidative, microbial, bacterial and sensory properties of minced beef. Journal of Food Measurement and Characterization. 14:3333-3344.
- Shaltout FA, Ibrahim HM. (1997). Quality evaluation of luncheon and Alexandrian sausage. Benha Vet Med J. 10(1):1-10.
- 56. Haq M, Dutta PL, Sultana N, Rahman MA. (2013). Production and quality assessment of fish burger from the grass carp, Ctenopharyngodon idella (Cuvier and Valenciennes, 1844). Journal of Fisheries. 1(1):42-47.
- Hassanin FS, Hassan MA, Shaltout FA, Shawqy NA, Abd-Elhameed GA. (2017). Chemical criteria of chicken meat. Benha Vet Med J. 33(2):457-464.
- Shaltout FA, Nassif MZ, Shakran AM. (2014). Quality of battered and breaded chicken meat products. Glob J Agric Food Safety Sci. 1(2):283-299.
- Saad SM, Shaltout FA, Elroos NAA, El-nahas SB. (2019). Antimicrobial Effect of Some Essential Oils on Some Pathogenic Bacteria in Minced Meat. J Food Sci Nutr Res. 2(1):012-020.
- Saad SM, Shaltout FA, Elroos NAA, El-nahas SB. (2019). Incidence of Staphylococci and E. coli in Meat and Some Meat Products. EC Nutrition. 14(6).

- Saad SM, Hassanin FS, Shaltout FA, Nassif MZ, Seif M. (2019). Prevalence of Methicillin-Resistant Staphylococcus Aureus in Some Ready-to-Eat Meat Products. AJBSR. 4(6):460-464.
- 62. Gaafar R, Hassanin FS, Shaltout FA, Zaghloul M. (2019). Hygienic profile of some ready to eat meat product sandwiches sold in Benha city, Qalubiya Governorate, Egypt. Benha Vet Med J. 37(2):16-21.
- Gaafar R, Hassanin FS, Shaltout FA, Zaghloul M. (2019). Molecular detection of enterotoxigenic Staphylococcus aureus in some ready to eat meat-based sandwiches. Benha Vet Med J. 37(2):22-26.
- 64. Efenberger-Szmechtyk M, Nowak A, Czyzowska A. (2021). Plant extracts rich in polyphenols: antibacterial agents and natural preservatives for meat and meat products. Crit Rev Food Sci Nutr. 61(1):149-178.
- 65. Da Silveira SM, Luciano FB, Fronza N, Cunha A, Scheuermann GN, Vieira CRW. (2014). Chemical composition and antibacterial activity of Laurus nobilis essential oil towards foodborne pathogens and its application in fresh Tuscan sausage stored at 7 °C. LWT-Food Science and Technology. 59(1):86-93.
- Shaltout FA, Salem AM, Mahmoud AH, Abd Elraheem KA. (2013). Bacterial aspect of cooked meat and offal at street vendors level. Benha Vet Med J. 24(1):320-328.
- 67. Shaltout FA, et al. (2012). Improvement of microbiological status of oriental sausage. JEVMA. 72(2):157-167.
- 68. Rubel SA, Yu ZN, Murshed HM, Islam SMA, Sultana D, Rahman SME, et al. (2020). Addition of olive (olea europaea) leaf extract as a source of natural antioxidant in mutton meatball stored at refrigeration temperature. Journal of Food Science and Technology. 58:1-9.
- Shaltout FA, Edris AM. (1999). Contamination of shawerma with pathogenic yeasts. Assiut Vet Med J. 40(64):34-39.