INTRODUCTION

Nowadays cancer is one of the main cause of death in around the world so that, in each year it is responsible for more 13% of death incidence [1]. The World Health Organization in 2015 estimated that 84 million people were died by cancer in the last decade [2]. In addition, GLOBOCAN 2012 reported 14.1 million new cancer per year in the world and estimated that by 2025 this number will raise up to 19.3 million [3]. Currently, four conventional therapeutic strategies for cancer therapy are surgery, radiation, chemotherapy and immunotherapy. Among these, chemotherapy is more common used but it has many disadvantages like high toxicity and side effects of anticancer drugs on healthy cells [4, 5].

Doxorubicin (DOX) is one of the most commonly used and most effective therapeutic anticancer drugs that has a high potency against of various cancers. In the other hand, DOX like the other anticancer drugs has severe side effects such as gastrointestinal disturbances, nausea, vomiting, hair loss, bone-marrow aplasia and congestive heart failure [6]. As regards to the importance of this matter, there is an urgent need to find the new strategies and materials for targeted drug delivery. In targeted drug delivery systems (TDDSs), suitable carriers can be utilized to protect anticancer drugs from premature degradation in body and also minimize their side effects on normal cells [7]. In this regard, so far a vast range of materials have been used as carrier in TDDSs for cancer therapy, such as polymeric nanoparticles and vesicles [3, 8], inorganic nanoparticles and vesicles [3, 8], inorganic materials [9, 10], liposomes [11], micelles [12] and so on.
Among proposed carriers, graphene oxide (GO) have received increasing attention in TDDSs due to its intrinsic advantages such as high surface area for physical adsorption of drugs (through π-π stacking), non-toxicity, high levels of cellular uptake, effective transportation capability and pH-dependent drug release behavior [13-16]. In addition, GO has numerous oxygen-containing functional groups like carboxyl groups, hydroxyl groups and epoxy which make it dispersible in physiological media [17]. These exclusive properties have made GO an ideal candidate for the targeted cellular delivery of anticancer drugs [18, 19].

Generally, a potent drug delivery system in cancer therapy must have the ability to deliver drugs to the target tissue while decreasing their side effects on normal tissues and also it must prevent premature degradation and perform a controlled release at the tumor site in response to a particular stimulus [7, 20]. For this purpose, to achieve the targeted delivery, GO could be functionalized with targeting ligands and in this way it could be quickly guided toward the tumor site [21]. Also, to prevent the premature release of drug before reaching to the target lesion, drug-loaded GO could be coated by a suitable coating.

In this study to deliver of doxorubicin, we used both of the targeting agent and appropriate coating in a drug delivery system, to targeted delivery and prevent the premature release of drug, respectively. In the first step, we modified GO with Fe₃O₄ magnetic nanoparticles by chemical co-precipitation method and then DOX loaded on it through π-π stacking interactions [22]. Hence, the GO-Fe₃O₄/DOX magnetic composite was prepared for magnetic targeted delivery of the cancer chemotherapy drug. In the second step, to prevent the premature release of drug during the guidance of composite toward the target tissue we utilized a technique which has been introduced in our previous works. In these works we successfully used titanium dioxide as a porous inorganic coating, to decrease the release of various drugs in controlled drug delivery systems [23-25]. In continuous of our previous studies and expanding this technique, we used it along with a targeting aspect in a drug delivery system for cancer therapy. Therefore, GO-Fe₃O₄/DOX composite was coated with TiO₂ by titanium tetra n-butoxide, as precursor and in this way GO-Fe₃O₄/DOX/TiO₂ nanohybrid organic-inorganic composite was synthesized. Then, the in vitro drug release of synthesized nanohybrid composite was investigated by UV-Vis absorbance in simulated buffer solutions.

**EXPERIMENTAL**

**Materials**

Powder flake Graphite (purity > 99.9%), ferric chloride hexa-hydrate (FeCl₃·6H₂O), ferrous sulfate hepta-hydrate (FeSO₄·7H₂O), NaNO₃, H₂SO₄, NH₃ and n-hexane were purchased from Merck. Doxorubicin (purity > 98%) was supplied from Exir pharmaceutical Co. of Iran. Titanium (IV) n-butoxide, Ti(O-nBu)₄ (97%), was obtained fromSigma–Aldrich. All of reagents and solvents were analytical grade and used without further purification. Human breast cancer cell line (MCF-7 cells) and human cervical cancer cell line (Hela cells) were supplied from pasture institute (Tehran, Iran).

**Characterization techniques**

UV-Vis absorbance (UV-2100, UNICO Instrument Corp.) was utilized to quantify the content of drug in buffer solution. FTIR spectra were recorded by FTIR spectrophotometer (Jasco 680-plus) in the range of 400-4000 cm⁻¹. To observe the morphology of coated and uncoated samples, a field emission scanning electron microscopy (FE-SEM) (HITACHI S-4160) in acceleration voltage of 15.0 kV was performed. Differential scanning calorimetry (DSC) of the samples were performed at 10 °C/ min in the 30-400 °C temperature range (BÄHR-Thermoanalyse GmbH – DSC 302). The size distribution and zeta potential of the samples were detected by dynamic light scattering using a Zeta sizer (Ver 6.00, Malvern Instrument Ltd., UK) by diluting the samples to 0.1 mg/mL in deionized (DI) water at 25 °C. Surface area and pore size distribution of samples were measured by the BET equation and BJH method, respectively [26, 27]. Before the adsorption–desorption measurements, the sample was degassed in vacuum at 100 °C for 6 h.

**Synthesis of graphene oxide**

Graphene oxide was synthesized by using an improved Hummers’ method according to the following procedure: in the first step, 0.5 g graphene powder and 0.25 g NaNO₃ were added to 12 mL sulfuric acid and vigorously stirred at 0°C for 10 h and then 1.5 g KMnO₄, oxidative agent, was added dropwise to the reaction mixture. The reaction temperature was increased to 45 °C and stirred for another 1 h. In the second step, 25 mL DI water was added to the reaction mixture and heated to 98 °C for 30 min, followed by cooling down to 0 °C for 1 h. Finally, to complete the reaction, 100 mL DI water and 4 mL H₂O₂ were added and reacted for 1 h. The obtained mixture was centrifuged and sequentially washed with 5% HCl solution (three times), and then washed several times with distilled water until the pH of the supernatant become neutral.

The obtained GO was sonicated for 30 min, filtered by filter paper and dried.

**Preparation of magnetic graphene oxide (GO-Fe304)**

In this section, through chemical co-precipitation of Fe₃O₄ magnetic nanoparticles on GO platelets, the magnetic GO was prepared. For this purpose, 50 mg GO was dispersed in 100 mL
DI water under nitrogen atmosphere. Then, 64 mg FeCl$_3$·6H$_2$O and excess amount of FeSO$_4$·7H$_2$O were added to the mixture. Reaction temperature was increased to 70 °C and reacted for 1 h under nitrogen atmosphere. Finally, a solution of NH$_3$·H$_2$O (2M) was added dropwise to the reaction mixture and stirred overnight. The GO–Fe$_3$O$_4$ product was obtained by magnetic separation and washed with water and dried.

**Preparation of GO-Fe$_3$O$_4$/DOX composites**

In this regard, initially 0.1 g GO-Fe$_3$O$_4$ was added to 50 mL DI water and it was sonicated for 30 min to be dispersed completely. Then, different amounts of DOX (10, 20, 30 and 40 wt% relative to the GO-Fe$_3$O$_4$) were added to the suspension and sonicated for another 30 min. After 24 h vigorous stirring, the obtained composites were dried using rotary evaporator at 70 °C. The information of prepared composites are summarized in Table 1.

**Kinetic and mechanism of drug release**

In this section, to investigate the in vitro drug release behavior of the prepared hybrid composites, they were studied kinetically using various mathematical models such as zero order, first order, Higuchi, Weibull-Baker-Lonsdale and Korsmeyer–Peppas. The Baker-Lonsdale model has been described for microcapsules or microspheres according to the equation (1):

$$ f_t = \frac{1}{2} \left[ 1 - \left( \frac{M_t}{M_w} \right)^{\frac{1}{n}} \right] \frac{M_w}{M_w} = K_i $$

In this equation, $M_t/M_w$ is a fraction of drug released at time $t$ and $k$ is the release rate constant which corresponds to the slope. To study the release kinetics, data obtained from in vitro drug release studies were plotted as [d(M/M$_w$)]/dt with respect to the root of time inverse [28].

**Cytotoxicity and cell viability assays**

The in vitro cytotoxicity of GO-Fe$_3$O$_4$/DOX/TiO$_2$ hybrid composite (B2) were investigated with MTT assay. For this purpose, MCF-7 and Hela cells were cultured in complete RPMI-1640 (Gibco, Scotland) medium supplemented with 10% fetal bovine serum and penicillin/streptomycin (100 units/ml). 200 μL of cell suspension (at concentration of 4×10$^4$ cells/well) was added and incubated for 24 h at 37 °C in a humidified atmosphere of 5% CO$_2$. Then, serial dilutions of B2 (20 μL; final concentration: 0.5, 1, 2 and 4 μM) were added and incubated for another 48 h. In the last step, to evaluate the cell viability, 20 μL of MTT solution (0.5% w/v) was added followed by incubation for 4 h. Then the media was replaced with 200 μL of DMSO and the plate shaken for 20 min at 37 °C. Absorbance was quantified at 540 nm by an ELISA plate reader (Awareness, USA). Each experiment included six wells for each condition and results expressed as mean ± standard deviation.

**RESULTS AND DISCUSSION**

**FTIR spectroscopy analysis**

(Figure 1) represents the FTIR spectra of the materials obtained at each step of the synthesis. The FTIR spectra of GO, GO-Fe$_3$O$_4$ and GO-Fe$_3$O$_4$/DOX composites (A1-A4) are given in the Figure. 1a. In the spectrum of GO, the presence of the oxygen functionalities on GO was confirmed by different ab-

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**Table 1**: Feed composition and drug content of GO-Fe$_3$O$_4$/DOX composites.

<table>
<thead>
<tr>
<th>Entry</th>
<th>Abbreviation</th>
<th>GO-Fe$_3$O$_4$ (g)</th>
<th>DOX (g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>GO-Fe$_3$O$_4$/DOX (10%)</td>
<td>A1</td>
<td>0.1</td>
<td>0.01</td>
</tr>
<tr>
<td>GO-Fe$_3$O$_4$/DOX (20%)</td>
<td>A2</td>
<td>0.1</td>
<td>0.02</td>
</tr>
<tr>
<td>GO-Fe$_3$O$_4$/DOX (30%)</td>
<td>A3</td>
<td>0.1</td>
<td>0.03</td>
</tr>
<tr>
<td>GO-Fe$_3$O$_4$/DOX (40%)</td>
<td>A4</td>
<td>0.1</td>
<td>0.04</td>
</tr>
</tbody>
</table>

**Table 2**: Different amounts of TiO$_2$ coated on GO-Fe$_3$O$_4$/DOX composites.

<table>
<thead>
<tr>
<th>Entry</th>
<th>Abbreviation</th>
<th>GO-Fe$_3$O$_4$/DOX</th>
<th>TiO$_2$ (g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>GO-Fe$_3$O$_4$/DOX/TiO$_2$ (40%, 30%)</td>
<td>B1</td>
<td>0.1</td>
<td>0.03</td>
</tr>
<tr>
<td>GO-Fe$_3$O$_4$/DOX/TiO$_2$ (40%, 40%)</td>
<td>B2</td>
<td>0.1</td>
<td>0.04</td>
</tr>
<tr>
<td>GO-Fe$_3$O$_4$/TiO$_2$ (40%, 50%)</td>
<td>B3</td>
<td>0.1</td>
<td>0.05</td>
</tr>
<tr>
<td>GO-Fe$_3$O$_4$/TiO$_2$ (40%, 60%)</td>
<td>B4</td>
<td>0.1</td>
<td>0.06</td>
</tr>
</tbody>
</table>

**Drug release studies**

In order to simulate the biological condition of body, the drug release of composites was investigated at the physiological temperature of 37 °C and pH 7.4. Moreover, for establishing environment similar to the cancer cells, we used a buffer solution with pH value of 5.4. These buffer solutions were prepared according to the same procedure in our previous works [23, 24]. The content of DOX in buffer solutions was quantified using UV-Vis absorbance at $\lambda_{max} = 490$ nm.
The FTIR spectra of (a) GO, GO-Fe\(_3\)O\(_4\), DOX, A1, A2, A3, A4; (b) TiO\(_2\), B1, B2, B3, B4.}

The FTIR spectra of GO-Fe\(_3\)O\(_4\)/DOX composites (A1-A4) and slightly shifted to the lower wavenumbers. Moreover, it is observed that by increasing the DOX content from A1 to A4 (10-40%) the intensity of these bonds is increased. These results confirm that DOX was successfully loaded onto GO-Fe\(_3\)O\(_4\). The FTIR spectra of GO-Fe\(_3\)O\(_4\)/DOX /TiO\(_2\) hybrid composites (B1-B4) are shown in (Figure 1b). For better comparison and understanding, the FTIR spectrum of amorphous TiO\(_2\) was added to the figure. According to the observed characteristic bonds for TiO\(_2\), the broad bonds appearing at 450–600 cm\(^{-1}\) in these spectra can be attributed to the vibration of the Ti-O-Ti bonds [23]. As can be seen, by increasing the TiO\(_2\) amount from B1 to B4, the intensity of this broad bond is increased.

**Figure 1:** FT-IR spectra of (a) GO, GO-Fe3O4, DOX, A1, A2, A3, A4; (b) TiO\(_2\), B1, B2, B3, B4.

### Differential scanning calorimetry (DSC)

The DSC thermograms were plotted simultaneously to compare the thermal properties of DOX, GO-Fe\(_3\)O\(_4\), GO-Fe\(_3\)O\(_4\)/DOX (40%) composite (A4) and GO-Fe\(_3\)O\(_4\)/DOX/TiO\(_2\) (40%, 50%) hybrid composite (B3) in Figure 2. For DSC of pure drug, DOX from the injection vial was first freeze-dried to obtain a dry powder. As can be seen, the thermogram of DOX exhibits a sharp endothermic peak at 218°C due to melting of the DOX (\(T_m = 218 \, ^\circC\)) [33]. In the case of GO-Fe\(_3\)O\(_4\), there is no peak in the range of used temperature, indicating that it has good thermal stability. In the thermogram of GO-Fe\(_3\)O\(_4\)/DOX (40%) composite (A4), the absence of a characteristic melting peak of DOX between 150 and 260°C confirms that no pure drug was precipitated during composition and drug was uniformly dispersed in the texture of composite [34]. However, in the higher temperatures a low endothermic peak at 283 °C was appeared that it can be attributed to decomposition of A4 composite. In the thermogram of the GO-Fe\(_3\)O\(_4\)/DOX/TiO\(_2\) (40%, 50%) hybrid composite (B3), a low endothermic peak at the higher temperature (332°C) was appeared that it shows the thermal stability of the composite was increased by coating with TiO\(_2\) compared to the uncoated composite [24]. These results could mean that the proposed method of gathering of the drug, matrix and inorganic coating has resulted to a uniform dispersion of the DOX in the GO-Fe\(_3\)O\(_4\) matrix with TiO\(_2\) coating.

**Figure 2:** DSC thermograms of DOX, GO-Fe3O4, A4 and B3.

### Zeta potential and DLS measurements

To measure the stability of the samples in suspension state and to determine their size distribution, we used the zeta potential and DLS analysis, respectively and results given in Figure 3. For this purpose, zeta potential of the GO, GO-Fe\(_3\)O\(_4\), A4 and B3 were measured by diluting the samples to 0.1 mg/mL in DI water without any additive at room temperature. As can be seen in the Figure. 3a, the zeta potential of GO illustrated a negative value (-45 mV) due to the abundance of OH and COOH groups. Compared to the GO, due to the positive charge of Fe\(_3\)O\(_4\) magnetic nanoparticles, the zeta potential of GO-Fe\(_3\)O\(_4\) increased slightly to -40 mV. In the case of A4 composite, the zeta potential again decreased to -51 mV due to the existence of OH and COOH groups in the structure of DOX. Compared to A4, the zeta potential of B3 hybrid composite finally decreased to -58 mV due to abundance of OH groups on the surface of TiO\(_2\). Since a zeta potential of greater than ± 30 mV is considered sufficient for stabilisation of a colloidal suspension [35], so it can be concluded that prepared hybrid composite (B3) is stable in water. In the other words, B3 does not tend to coagulate in the water and so it can be utilized as...
a good carrier in TDDSs. Moreover, for particle size determination of the samples, we applied the dynamic light scattering (DLS) analysis with experimental conditions similar to the zeta potential analysis and data given in the Fig. 3b. The z-average values for GO, GO-Fe₃O₄, A4 and B3 were obtained 92, 95, 115 and 126 nm, respectively.

**Figure 3:** Zeta potential (a) and size distribution (b) of GO, GO-Fe₃O₄, A4 and B3.

**FE-SEM images**

In order to observe the morphology of the optimum GO-Fe₃O₄/DOX composite (A4), before and after coating by amorphous TiO₂, we used field-emission scanning electron microscopy (FE-SEM) analysis. The FE-SEM image of the GO-Fe₃O₄/DOX composite (A4) (Figure 4a,b) revealed a sheet-like structure with different orientations and accumulated morphology [36]. According to the DSC thermogram of A4 composite we can judged that DOX is dispersed in the GO-Fe₃O₄ matrix, so it can be concluded that the presence of a large number of oxygen-containing functional groups on the surface of GO and DOX, probably is the main factor of this accumulation [37]. After coating A4 composite with a porous TiO₂, the morphology of the GO-Fe₃O₄/DOX/TiO₂ hybrid composite (B3) presented a uniform coating of porous TiO₂ on the surface of A4 composite (Figure 4c,d) [24,38]. It could be expected that by formation a porous shell of TiO₂ around A4 composite, the premature release of the DOX from GO-Fe₃O₄ matrix would be reduced.

**Figure 4:** FESEM images of A4 (a,b) and B3 (c,d).

**Drug release studies**

The DOX release tests were studied by suspending the prepared composites in simulated buffer solutions at 37 °C. For this purpose, the drug release profile was tested at pH 7.4 and 5.4, which correspond to the physiological pH and endosomal pH of cancer cell, respectively [42]. The release was monitored with UV-Vis spectrophotometry by observing the change in absorbance of the characteristic band of DOX at 500 nm. All experiments were done in triplicate and the results were averaged. In the first step, we examined the drug release of the GO-Fe₃O₄/DOX composites with 10, 20, 30 and 40 wt% of DOX (A1-A4) (Table 1). During the preparation of GO-Fe₃O₄/DOX composites, we expected that DOX adheres onto the surface of GO via strong π-π stacking interactions. In addition, the NH₂ group of DOX is expected to form additional hydrogen bindings with OH groups of the GO. Figure 6a represents the release profiles of DOX from the GO-Fe₃O₄/DOX composites in pH 7.4. As can be seen, there was an acceptable sustained release of the drug from all of the composites for 54 h, but dramatically they released about >30% of drug in the first 1 h and >40% after 3 h which it consider a negative point for a TDDS. Nevertheless, as we demonstrated in pre-
Previous studies, this problem can be solved by formation of a porous inorganic coating like TiO$_2$ around composites. Therefore in the second step, the composite containing 40 wt% of DOX (A4) (with highest correlation coefficient) was selected as optimum formulation and was coated by 50 wt% of TiO$_2$ (Scheme 1).

**Scheme 1:** Schematic presentation of GO-Fe$_3$O$_4$/DOX/TiO$_2$ hybrid composite formation mechanism.

In this way, according to the method mentioned in section 2.4, the Fe$_3$O$_4$/DOX/TiO$_2$ (40%, 50%) hybrid composite (B3) was synthesized and drug release was examined once again. As can be observed in the Figure 6b, the result was as expected and the coated composite (B3) released only 3% of DOX in the first 1 h and 6% after 2 h and also whole of drug was released after 120 h (data not shown). It is likely that TiO$_2$ coating remains strongly on the composite through the strong hydrogen bonding interactions between composite and OH groups on the surface of TiO$_2$. Moreover, the formation of covalent bonds between the composite and TiO$_2$ (during the hydrolysis of titanium tetra butoxide) could not be certainly rejected. In continuous, to investigate the effect of TiO$_2$ amount on the drug release, the optimum composite (A4) was coated with different amount of TiO$_2$. Hence, other GO-Fe$_3$O$_4$/DOX/TiO$_2$ hybrid composites with different amount of TiO$_2$ were prepared (B1, B2 and B4) and their information summarized in Table 2. The drug release profiles of the B1, B2 and B4 composites were added to the Fig. 6b for better comparison. As is clear, for B1 and B2 with 30 and 40 wt% of TiO$_2$ coating respectively, the amount of released DOX is increased compared to the B3 (with 50 wt% TiO$_2$) while the performance of B4 (60 wt% TiO$_2$) was about similar to B3. Based on the better performance (compared to B1 and B2) and lower amount of TiO$_2$ (compared to B4), the GO-Fe$_3$O$_4$/DOX/TiO$_2$ (40%, 50%) hybrid composite (B3) was selected as the optimum composite. In the last step, as the endosomal pH of cancer cells is slightly acidic, we used the buffer solution with pH of 5.4 to study the effect of pH on drug release. The results revealed that the release of DOX from B3 is pH-dependent. As can be seen in the Figure 6c, in slightly acidic solutions (pH 5.4) simulated to the reduced pH micro-environment typical of the cancerous cells, the release rate is significantly enhanced. In the pH 5.4, the amount of DOX released after 80 h is about 75% while in the pH 7.4 it was 15%. This difference may be caused by weakening of hydrogen bonds between the DOX and GO. Under acidic condition at pH 5.4, the NH$_2$ group of DOX is protonated and H$^+$ in solution would compete with the hydrogen bond-forming group and weaken the hydrogen bond interactions, as a result the DOX would faster release from composite.

Generally, the obtained results indicates that the GO-Fe$_3$O$_4$/DOX/TiO$_2$ hybrid composite could be used as useful carrier in TDDS, because the pH-sensitive drug release could facilitate hybrid composite to release its drug cargo in the acidic (pH < 6) endosomal environment after cellular uptake and increase the cytotoxicity toward cancer cells, while avoid injurious side effects in normal tissue.

The in vitro drug release data from various GO-Fe$_3$O$_4$/DOX/TiO$_2$ hybrid composites (B1-B4) were investigated kinetically using various mathematical models. According to the obtained R$^2$ values of the curves for various mathematical models, it was shown that there is a very good fitting between experimental data and Baker-Lonsdale model.
Cell viability assays

Figure 7a presents the viability of MCF-7 and Hela cancerous cells after treatment with different concentrations of GO-Fe$_3$O$_4$/DOX/TiO$_2$ hybrid composite (B3) after 48 h incubation time. When MCF-7 cells were treated with B3, they generally showed lower viability compared to Hela cells. As can be seen, different concentrations of B3 (0.5-4 μM) reduced viability of MCF-7 and Hela cells down to 41% and 26% respectively, which confirmed its superior responsive to cancer cell lines. Also, the cytotoxicity of GO-Fe$_3$O$_4$/TiO$_2$ composite (DOX free) against these cells with experimental conditions similar to the B3 was explored and results given in the Figure. 7b. As is clear, the DOX free composite shows very low cytotoxic effect on cancer cells which confirms its biocompatible and inert nature.

Figure 7: Cell viability of MCF-7 and Hela cells exposed to different concentrations of B3 (a) and GO-Fe$_3$O$_4$/TiO$_2$ (DOX free) (b) composites after 48 h incubation.

ACKNOWLEDGMENTS

Thanks are due to the Research Council of Isfahan University of Technology and Center of Excellency in the Chemistry Department of Isfahan University of Technology for supporting this work.

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